Supplementary Information

Explaining the striking difference in twist-stretch coupling between DNA and RNA: A comparative molecular dynamics analysis

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Table S1. Overview on experimental measurements of the twist-stretch coupling for double-stranded DNA and double-stranded RNA.

Measurement	Twist stretch coupling:	Twist-stretch
	change in length per	coupling
	change in twist	parameter D
	(Å/deg)	(unitless)
Double-stranded DNA; rotor bead assay [1]	0.014 ± 0.003	-22 ± 4.9
Double-stranded DNA; magnetic tweezers [2]	0.012 ± 0.006	−17 ± 7
Double-stranded DNA; optical torque wrench [3]	N.A.	−21 ± 1
Double-stranded DNA; magnetic tweezers [4]	0.012 ± 0.003	−17 ± 5
Double-stranded RNA;magnetic tweezers [4]	-0.024 ± 0.001	11.5 ± 3.3

- 1. Gore, J., Bryant, Z., Nollmann, M., Le, M.U., Cozzarelli, N.R. and Bustamante, C. (2006) DNA overwinds when stretched. *Nature*, **442**, 836-839.
- 2. Lionnet, T., Joubaud, S., Lavery, R., Bensimon, D. and Croquette, V. (2006) Wringing out DNA. *Phys Rev Lett*, **96**, 178102.
- 3. Sheinin, M.Y. and Wang, M.D. (2009) Twist-stretch coupling and phase transition during DNA supercoiling. *Phys Chem Chem Phys*, **11**, 4800-4803.
- 4. Lipfert, J., Skinner, G.M., Keegstra, J.M., Hensgens, T., Jager, T., Dulin, D., Kober, M., Yu, Z., Donkers, S.P., Chou, F.C. *et al.* (2014) Double-stranded RNA under force and torque: Similarities to and striking differences from double-stranded DNA. *Proc Natl Acad Sci U S A*, **111**, 15408-15413.

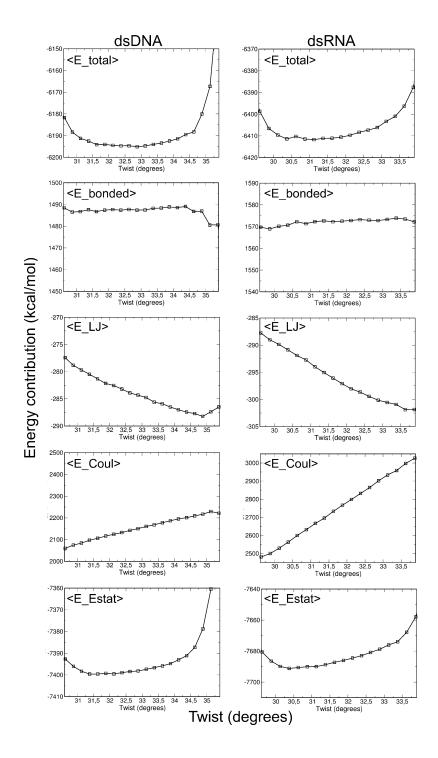


Figure S1. Average energy contributions vs. mean twist of sampled states during 1 µs unrestraint simulations of DNA and RNA. Snapshots were evaluated using the MMGBSA approach (see Methods). The mean total energy (<E_total>) is the sum of all bonded contributions (<E_bonded>) such as bond length, bond angle and dihedral contributions and Lennard-Jones (<E_LJ) as well as electrostatic contributions (<E_Estat>). The electrostatic contribution is the sum of Coulomb interactions (<E_Coul>) and the Generalized Born solvation term (not shown, represents the difference between E_Estat and E_Coul).

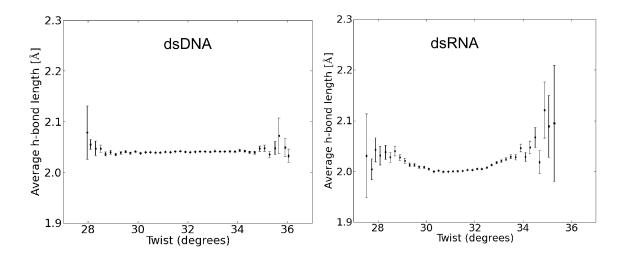


Figure S2. Correlation of the mean hydrogen bond length of Watson Crick base pairs vs. twist during unrestraint MD-simulations.

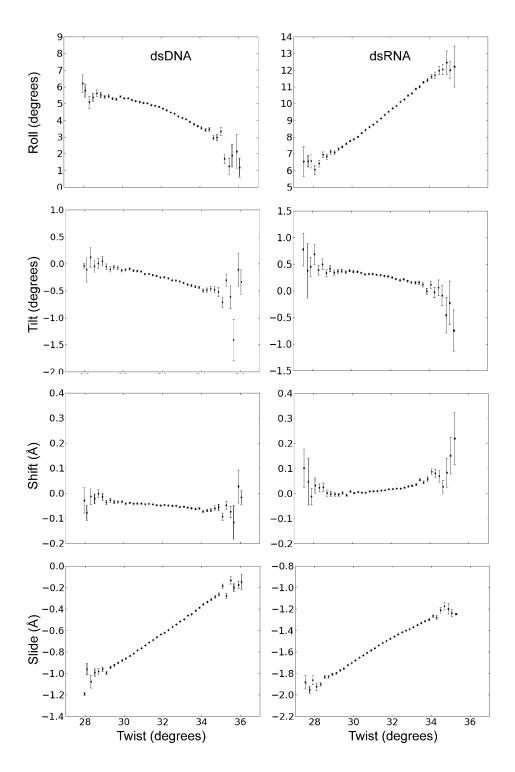


Figure S3. Correlation of local base pair step parameters with helical twist of DNA (left column of plots) and RNA (right column of plots). The plots were generated from the analysis of a total of ~100000 regularily spaced snapshots taken during ~1 μs unrestraint simulations. The data was analysed in twist intervals of 0.2° (x-axis) calculating the mean helical twist and the mean selected helical coordinate within the interval. Errors (shown as bars) were calculated as mean standard errors within each intervall.

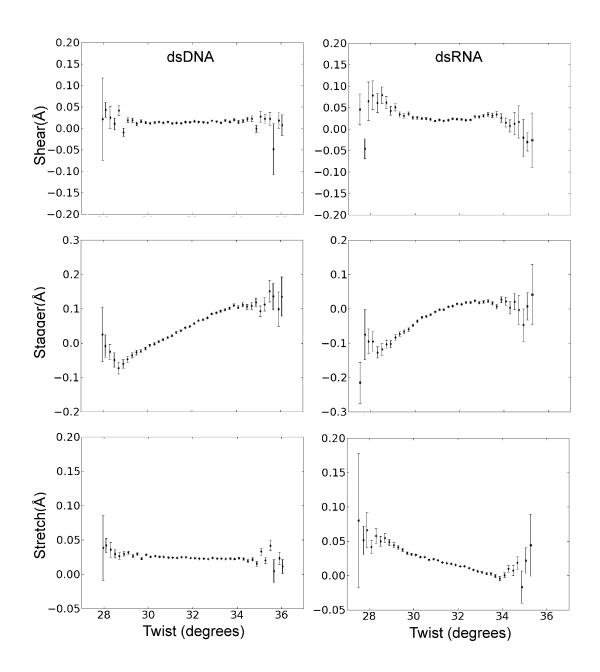


Figure S4. Correlation of translational local base pair parameters with helical twist of DNA (left column of plots) and RNA (right column of plots). The plots were generated from the analysis of a total of ~100000 regularily spaced snapshots taken during ~1 μs unrestraint simulations. The data was analysed in the same way as explained in the legend of Figure S1.

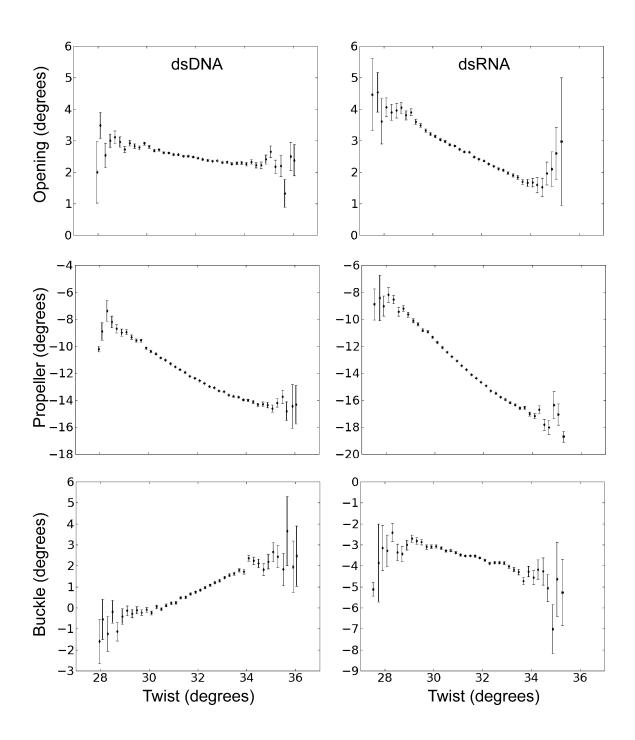


Figure S5. Correlation of angular local base pair parameters with helical twist of DNA (left column of plots) and RNA (right column of plots). The plots were generated from the analysis of a total of 100000 regularily spaced snapshots taken during 1 μ s unrestraint simulations. The data was analysed in same way as explained in the legend of Figure S1.

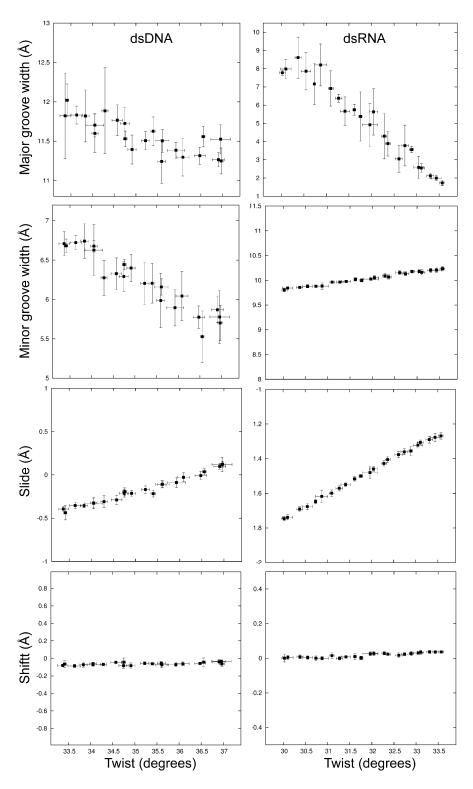


Figure S6. Variation of major and minor groove width, slide and shift during simulations with a torque restraint on the total twist of the DNA (left panels) and RNA (right panels). The mean twist per base pair step was changed in steps of ~0.15°. Error bars were calculated by splitting the recorded data for each twist restaining simulation into 5 intervals and calculating the standard variation over these intervals.

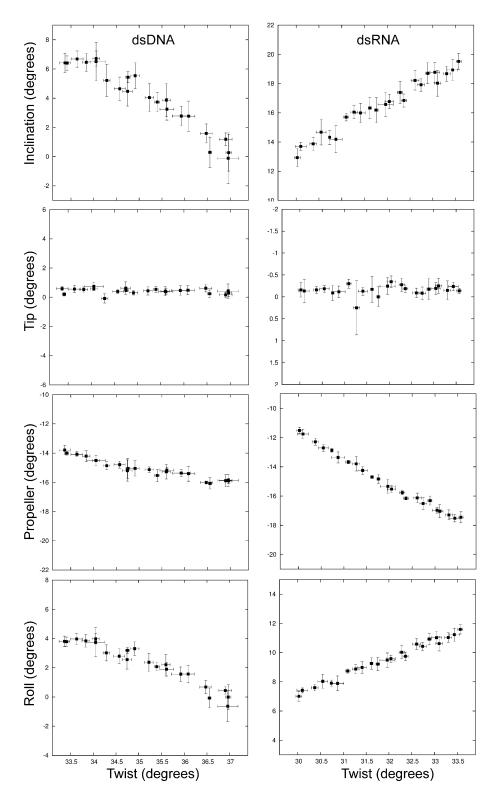


Figure S7. Variation of selected angular helical parameters inclination and tip, roll and propeller during simulations with a torque restraint on the total twist of the DNA (left panels) and RNA (right panels). The mean twist per base pair step was changed in steps of ~0.15°. Error bars were calculated by splitting the recorded data for each twist rrestaining simulation into 5 intervals and calculating the standard variation over these intervals.